

# **Development and deployment of the BioRAPTR 2.0 FRD Dispenser through collaboration**, and necessity, and evaluation of dispense and liquid handling technologies.

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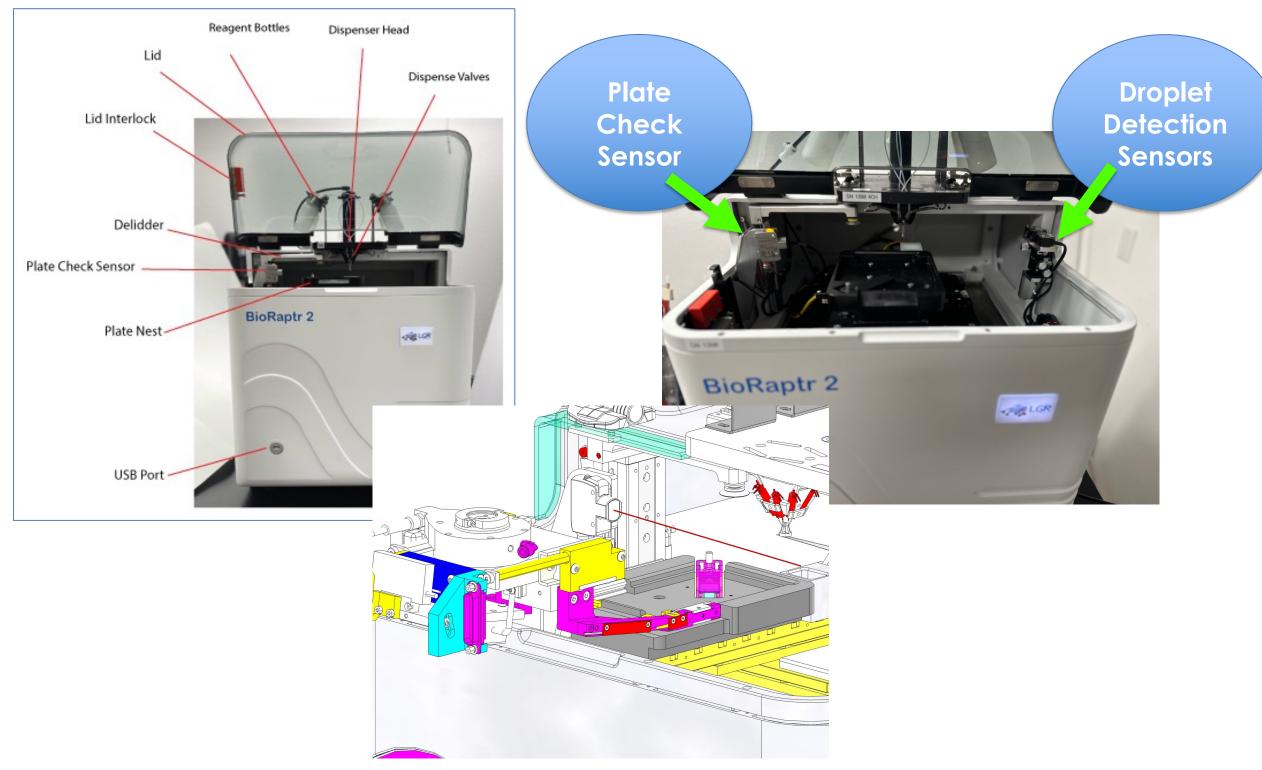
#### Introduction

The National Center For Advancing Translational Sciences (NCATS) has historically employed 1<sup>st</sup> generation BioRAPTR Flying Reagent Dispensers for offline assay validation/development and on our qHTS platforms. NCATS' fleet of 1<sup>st</sup> generation BioRAPTRs were at end-of-life due to unsupported hardware and software. The need to maintain continuity and retain the functionality of the 1st generation BioRAPTRs prompted NCATS to seek out a vendor to undertake significant revamp to these devices. Let's Go Robotics, Inc., in collaboration with NCATS, took on the task of retrofitting these 1<sup>st</sup> generation BioRAPTRs platforms with new internal electronics, controllers, software and GUI to create the BioRAPTR 2.0. Numerous software features were added beyond the 1<sup>st</sup> generation of BioRAPTRs. These include semiauto volume scaling, multi-step dispense protocols, user access management, true column dispense, auto-calculated gradient dispenses, multiple liquid class calibrations stored on-board, backward compatibility with existing Excel based dispense definitions, simplified plate definition control widget, robot gripper compatible plate nest, droplet detection, among others. LGR has also added plate detection sensors and reporting to prevent crashes involving the dispenser tips, a common occurrence with the 1st generation BioRAPTRs. NCATS collaborated with LGR to perform in-lab testing of the software interface, mechanical performance and validation of dispense volumes across various liquid classes and plate types. Extensive testing and resulting feedback from benchwork completed at NCATS by staff engineers and scientists provided to LGR's engineers and software developer further refined the hardware and software.

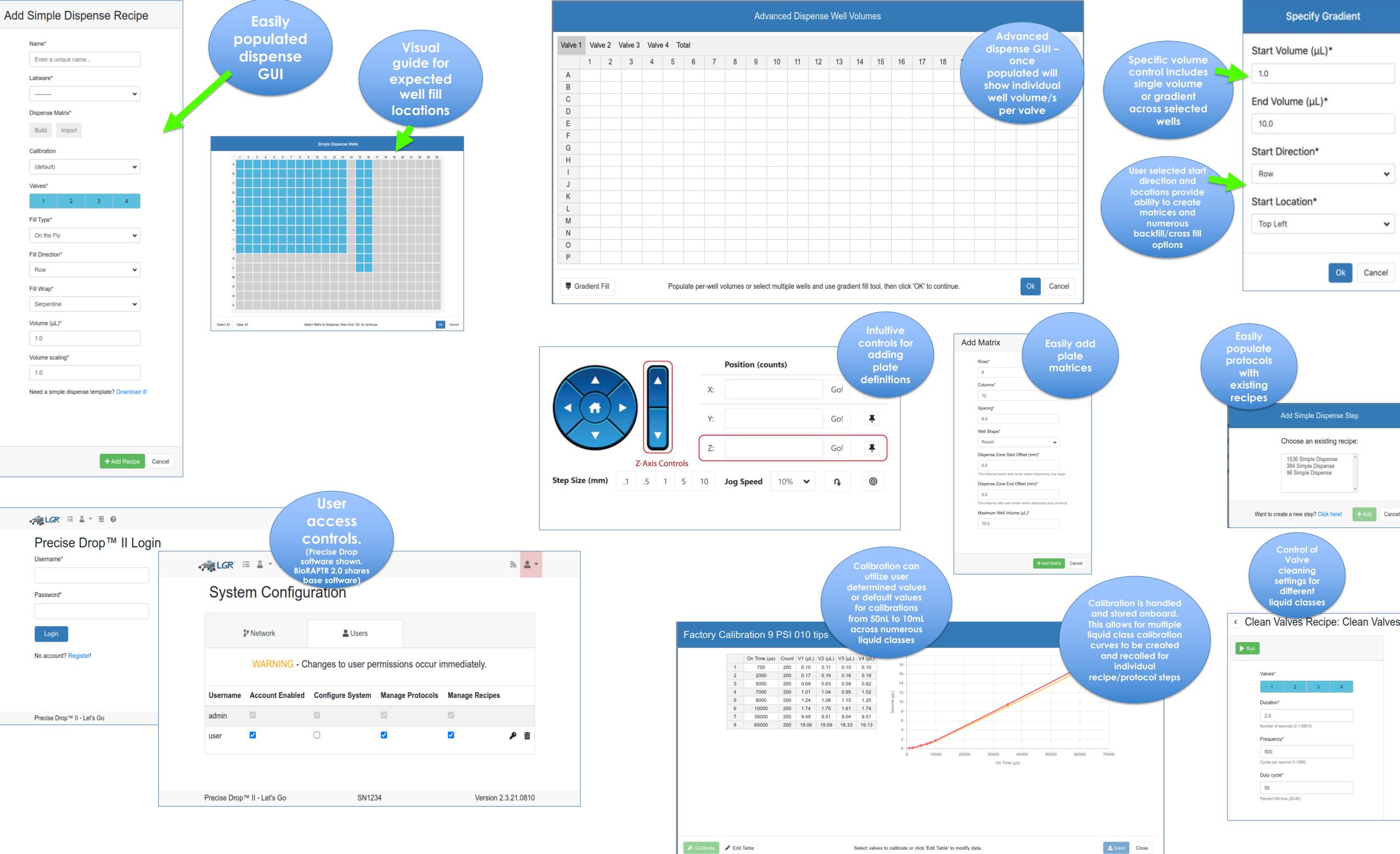
## Objective

Evaluate hardware, software and dispense volume accuracy of Let's Go Robotics, Inc. BioRAPTR 2.0

## **BioRAPTR 2.0 Overview**



#### **BioRAPTR 2.0 Software Overview**

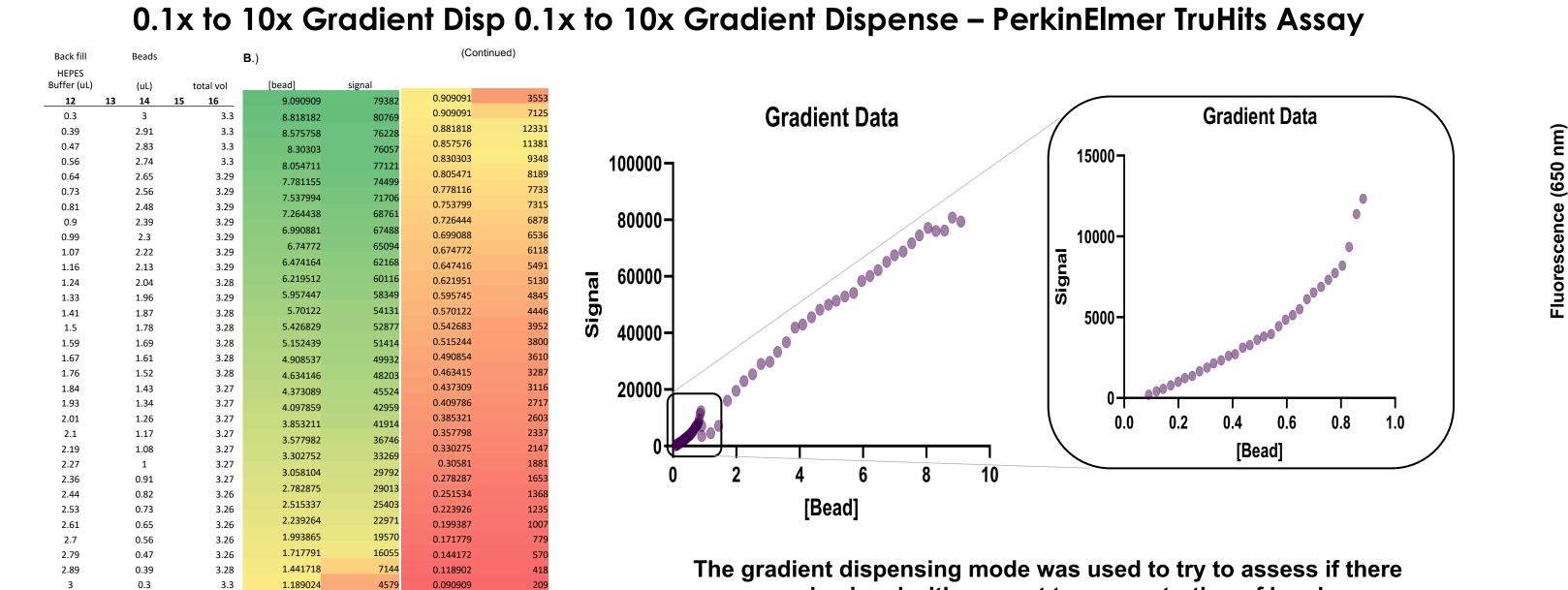


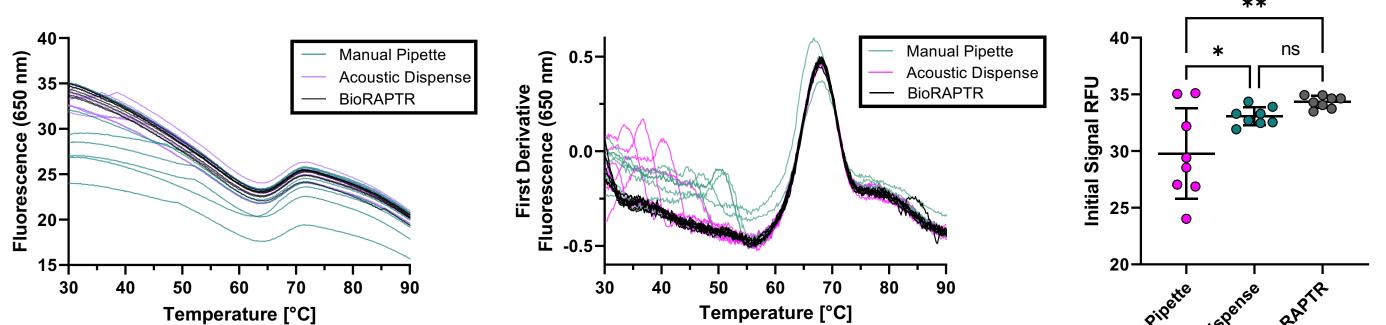
Existing devices are used to retrofit 1<sup>st</sup> generation BioRAPTRs to the LGR BioRAPTR 2.0 platform. New Z axis motors and internal automation control components are utilized to modernize these devices. The addition of plate detection and droplet detection sensors provides increased functionality and equipment protection. Also, alternative plate nests can be installed for the myriad robot grippers found on automation systems. LGR has removed the need for X Y stage end limit sensors, thus resolving issues related to valve leaks and/or other unintended liquid events. In the 1<sup>st</sup> generation BioRAPTR any liquid exposure to the X Y axis limit sensor would damage the stage, make it inoperable and result in costly repairs and device downtime

🚓 LGR 🗧	Bio R	ftware shown. APTR 2.0 shares		2
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# Examples of Gradient Dispense<sup>1</sup> and Comparative Dispense Accuracy<sup>2</sup>

Michael Ronzetti<sup>1</sup>





**Comparison of Dispense Methods for HIS-DSF** 

## Conclusions

- Retrofit of 1<sup>st</sup> generation BioRAPTRs to the LGR BioRAPTR 2.0 platform provides continuity of assay development/validations and qHTS assays performed at NCATS.
- Anecdotal user feedback of the software GUI has been extremely positive.
- Validation of accuracy of gradient dispenses (example<sup>1</sup>) shows consistent volumes dispensed per the onboard software calculations. This removes potential errors in user calculations when populating gradient dispenses.
- Validation of dispense volumes compared to manual and acoustic dispense methods (example<sup>2</sup>) show greater accuracy over manual dispenses and comparable dispenses using an acoustic dispenser vs.

was a cap in signal with respect to concentration of beads.

Two batches were dispensed; 0.1x to 1x and 1x to 10x concentrations.

Brooklyn Aragon<sup>1</sup>

Testing dispense methods using an established HIS-DSF assay against in-house protease target. (Left) Raw thermogram traces for n = 8 replicates of each liquid handling method dispense of 1 µL protein-dye mixture into 1536-well Roche PCR plates and tested in a standard DSF melting experiment. (Middle) First derivative of the n=8 replicate samples from each dispense method. (Right) Scatter plot of the sample initial fluorescence signal from each dispense method. Lines represent the means of the n=8 replicates with error bars representing the standard deviation with significance values derived from one-way ANOVA with Tukey's multiple comparisons between groups (\* = p<0.05, \*\* = p < 0.01).

the BioRAPTR 2.0. This dispense was made to a 1536 well Roche Light Cycler Plate. The acoustic dispense took 36 +/- minutes vs 1.5 minutes with the BioRAPTR 2.0. 1<sup>st</sup> generation BioRAPTRs could not accomplish this dispense due to plate well targeting issues - droplet release timing, location within the well, plate materials and well design.

Additionally, retrofit of existing 1<sup>st</sup> generation BioRAPTRs to the LGR **BioRAPTR 2.0 platform provided significant cost savings over** maintaining existing devices and/or acquiring, deploying and training staff to use new instrumentation.